



Overview

1. Fixed vs. Random
2. Pseudo-R²s
3. SEM Example of mixed models
4. Causal Modeling with Random Effects
5. Fully hierarchical SEM

Fixed vs. Random. Comparison

| Fixed | Random |
|---|--|
| Interested in drawing inferences / making predictions | Not particularly interested in any particular value or level |
| Represent values from the entire 'universe' of interest | A (random) sample from a larger pool of potential values |
| Levels not interchangeable | Levels interchangeable (could swap / relabel levels without any change in meaning) |
| Directly manipulated | Introduces incidental error (e.g., between subjects, blocks, sites, etc.) |
| Few levels / worth sacrificing d.f. to fit model | Many levels / cannot sacrifice d.f. to fit model |

Fixed vs. Random. Why mixed models?

- More power than modeling the means of groups
- Reduces degrees of freedom necessary to fit model and estimate parameters (vs. modeling as a fixed effect)
- Accounts for uneven sampling within groups by using information across groups to inform the individual group means
- Can account for *non-independence* of observations by explicitly modeling their covariances (e.g., among sites, individuals, etc.)

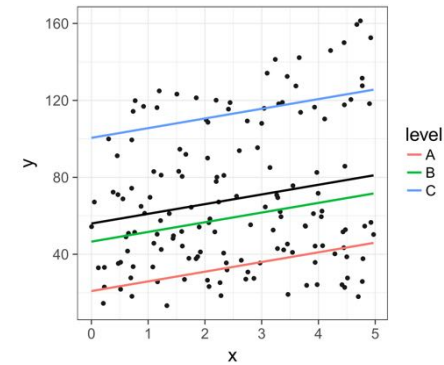
Fixed vs. Random. Random structure

Different configurations of random structure:

1. Varying intercept, fixed slope
2. Fixed intercept, varying slope
3. Varying intercept, varying slope

Fixed vs. Random. Varying intercept

- Estimates different intercept, same slope for all levels of the random effect

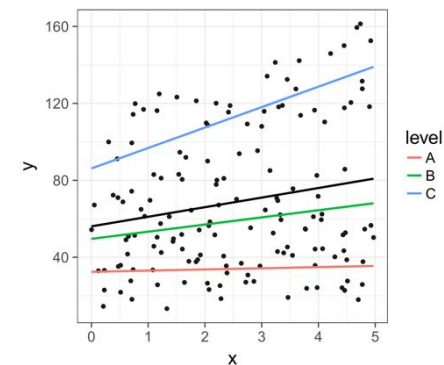


Fixed vs. Random. Varying intercept

- Good for block designs, repeated measures
- Can lead to overconfident estimates if levels are expected to respond differently (e.g., individuals in a drug trial)

Fixed vs. Random. Varying intercept AND slope

- Estimates different slope, different intercept for all levels



Fixed vs. Random. Varying intercept AND slope

- Addresses multiple sources of non-independence of within and between levels, leading to lower Type I *and* Type II error
- Random slopes can be extracted and used in other analyses (get error from lmerTools)
- Computationally intensive, may lead to non-convergence

Fixed vs. Random. Nesting

- Hierarchical models represent nested random terms (e.g., site within region)
- Nesting further addresses non-independence by modeling correlations within *and* between levels of the hierarchy
- Good for stratified sampling designs (varying intercept) and split-plot designs (varying slope, varying intercept)

Fixed vs. Random. Crossed effects

- Multiple random effects that are not nested but apply independently to the observation (e.g., space *and* time)

Fixed vs. Random. Random structures

| | |
|--|--|
| (1 group) | random group intercept |
| (x group) = (1+x group) | random slope of x within group with correlated intercept |
| (0+x group) = (-1+x group) | random slope of x within group: no variation in intercept |
| (1 group) + (0+x group) | uncorrelated random intercept and random slope within group |
| (1 site/block) = (1 site)+(1 site:block) | intercept varying among sites and among blocks within sites (nested random effects) |
| site+(1 site:block) | <i>fixed</i> effect of sites plus random variation in intercept among blocks within sites |
| (x site/block) = (x site)+(x site:block) = (1 + x site)+(1+x site:block) | slope and intercept varying among sites and among blocks within sites |
| (x1 site)+(x2 block) | two different effects, varying at different levels |
| x*site+(x site:block) | <i>fixed</i> effect variation of slope and intercept varying among sites and random variation of slope and intercept among blocks within sites |
| (1 group1)+(1 group2) | intercept varying among crossed random effects (e.g. site, year) |

<http://glmm.wikidot.com/faq>

Fixed vs. Random. A warning

- Assumes fixed and random effects are *uncorrelated*
 - e.g., all of your warm data points don't come from a different site than your cool data points
- If possible, fit random effects as fixed effects and compare parameter estimates of other predictors
- Need to ensure appropriate replication at *lowest* level of nested factors (5-6 levels, *minimum*) – otherwise, fit as fixed effects

Fixed vs. Random. Different distributions

- *lme4* can fit many kinds of different distributions using `glmer`
- Does not provide *P*-values (d.d.f uncertain, see: <https://stat.ethz.ch/pipermail/r-help/2006-May/094769.html>)
 - Need to turn to *pbkrtest* package which estimates d.d.f. using the Kenward-Rogers approximation (less finicky than *lmerTest*)
 - *piecewiseSEM* does this for you automatically using `coefs`

Fixed vs. Random. Different distributions

- *nlme* can only handle normal distributions
 - Ives (2015): "For testing the significance of regression coefficients, go ahead and log-transform count data"
- `glmmPQL` in the *MASS* package uses penalized quasi-likelihood to fit models, can incorporate many different distributions and their quasi- equivalents (e.g., quasi-Poisson)
 - Quasi-distributions estimate a separate term for how the variance scales with the mean, so ideal for over/under-dispersed data
 - Quasi-likelihood means no likelihood based statistics (e.g., AIC, LRT, etc.) for any models fit with `glmmPQL`
 - Implementing R^2 for quasi-distributions right now

Fixed vs. Random. Troubleshooting

- R has the most infuriating error messages
- Can sometimes solve by switching to a different optimizer
 - `lmeControl(opt = "optim")` usually works
- Reduce tolerance for convergence
 - `lmeControl(tol = 1e-4)`
- Respecify random structure
 - Optimizer constrained to have $cov > 0$, can sometimes get stuck bouncing around when random components are very close to 0
- <https://stackoverflow.com/>
 - Ben Bolker to the rescue! <https://dynamicecology.wordpress.com/2013/10/04/wwbbd/>

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Pseudo-R²s. Omnibus test

- Fisher's C is the global fit statistic for local estimation but has many shortcomings:
 - Sensitive to the number of d-sep tests and the complexity of the model (harder to reject as the complexity increases)
 - Sensitive to the size of the dataset (e.g., high n leads to low P)
 - Fails symmetry when dealing with unlinked non-normal intermediate variables

Pseudo-R²s. Local tests

- How do we infer the confidence in our SEM?
 - Examine standard errors of individual paths, qualitatively assess cumulative precision
 - Explore variance explained (i.e., R^2), qualitatively assess cumulative precision

Pseudo-R²s. General linear regression

- Coefficient of determination (R^2) = proportion of variance in response explained by fixed effects
- For OLS regression, simply 1- the ratio of unexplained (error) variance (e.g., SS_{error}) over the total explained variance (e.g., SS_{total})
- Ranges (0, 1), independent of sample size
- Not good for model comparisons since R^2 monotonically increases with model complexity

Pseudo-R²s. Generalized linear regression

- Likelihood estimation is not attempting to minimize variance but instead obtain parameters that maximize the likelihood of having observed the data
- In a likelihood framework, equivalent R² = 1- the ratio of the log-likelihood of the full model over the log-likelihood of the null (intercept-only) model
- Leads to identical R² as OLS for normal (Gaussian) distributions, not so for GLM – need to use likelihood-based pseudo-R² (e.g., McFadden, Nagelkerke)

Pseudo-R²s. Generalized mixed models

- Becomes even worse for mixed models because variance is partitioned among levels of the random factor, so what is the error variance?
- Need a new formulation of R² :
 - Marginal R² = variance explained by fixed effects only

$$R_{GLMM(m)}^2 = \frac{\sigma_f^2}{\sigma_f^2 + \sum_{l=1}^u \sigma_l^2 + \sigma_e^2 + \sigma_d^2}$$

Pseudo-R²s. Generalized mixed models

- Conditional R² = variance explained by both the fixed and random effects

$$R_{GLMM(c)}^2 = \frac{\sigma_f^2 + \sum_{l=1}^u \sigma_l^2}{\sigma_f^2 + \sum_{l=1}^u \sigma_l^2 + \sigma_e^2 + \sigma_d^2}$$

Pseudo-R²s. Generalized mixed models

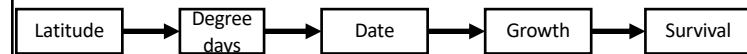
- Comparison of marginal and conditional R² can lead to roundabout assessment of ‘significance’ of the random effects (e.g., if conditional R² is larger relative to marginal R²)
- Best to report both and allow readers to determine how their magnitude affects the inferences

Overview

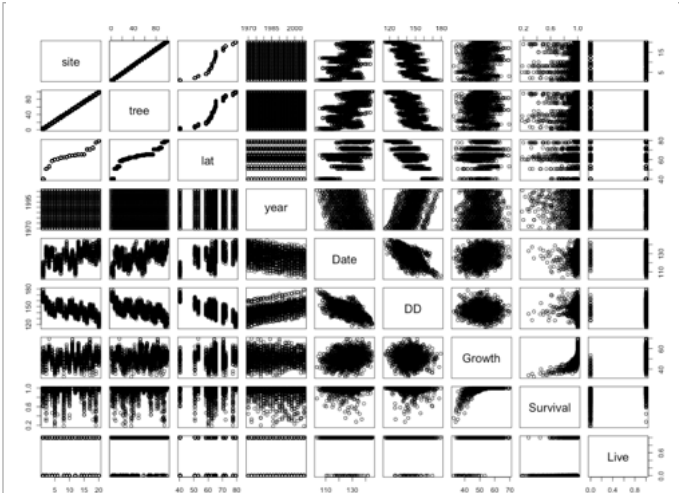
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SEM Example. Shipley 2009

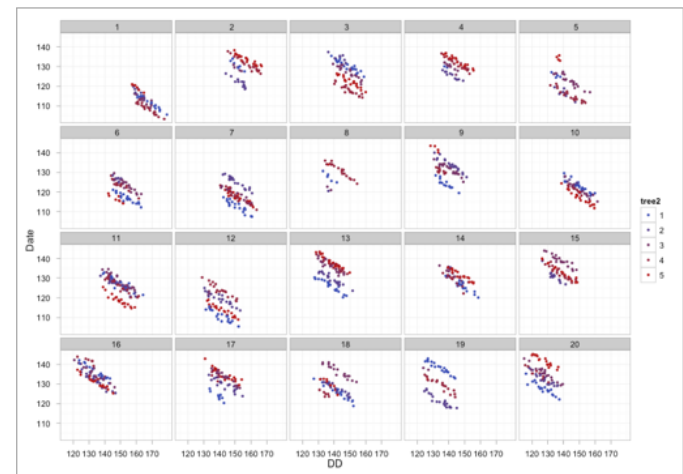
- Hypothetical dataset: predicting latitude effect on survival of a tree species
- Repeated measures on 5 subjects at 20 sites from 1970-2006
- Survival (0/1) influenced by phenology (degree days until bud break, Julian days until bud break), size (stem diameter growth)



The Simulated Data

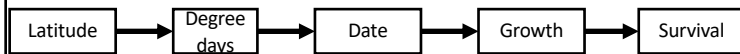


Nested Structure in the Data

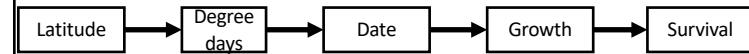


SEM Example. Shipley 2009

- Two distributions: normal, binary (survival)
- Random effects:
 - Site-only: latitude
 - Site and year: degree days, date
 - Site, year, and subject: diameter, survival

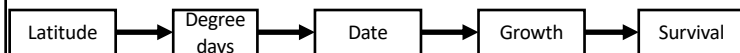


SEM Example. What is the basis set?



- Date \perp Lat | (Degree days)
- Growth \perp Lat | (Date)
- Survival \perp Lat | (Growth)
- Growth \perp Degree days | (Date, Lat)
- Survival \perp Degree days | (Growth, Lat)
- Survival \perp Date | (Growth, Degree days)

SEM Example. List of equations



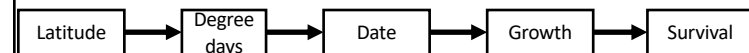
```

# Load data
shipley <- read.csv("shipley.csv")

# Create list of structural equations
shipley.sem <- psem(
  lme(DD ~ lat, random = ~1|site/tree, na.action = na.omit,
    data = shipley),
  lme(Date ~ DD, random = ~1|site/tree, na.action = na.omit,
    data = shipley),
  lme(Growth ~ Date, random = ~1|site/tree, na.action = na.omit,
    data = shipley),
  glmer(Live ~ Growth + (1|site) + (1|tree),
    family=binomial(link = "logit"), data = shipley)
)

# Get summary
summary(shipley.sem)
  
```

SEM Example. D-sep tests



Tests of directed separation:

| | Independ.Claim | Estimate | Std.Error | DF | Crit.Value | P.value |
|--------------------|----------------|----------|-----------|---------|------------|---------|
| Date ~ lat + ... | -0.0091 | 0.1135 | 18 | -0.0798 | 0.9373 | |
| Growth ~ lat + ... | -0.0989 | 0.1107 | 18 | -0.8929 | 0.3837 | |
| Live ~ lat + ... | 0.0305 | 0.0297 | NA | 1.0280 | 0.3039 | |
| Growth ~ DD + ... | -0.0106 | 0.0358 | 1329 | -0.2967 | 0.7667 | |
| Live ~ DD + ... | 0.0272 | 0.0271 | NA | 1.0038 | 0.3155 | |
| Live ~ Date + ... | -0.0466 | 0.0298 | NA | -1.5626 | 0.1181 | |

Global goodness-of-fit:

Fisher's C = 11.538 with P-value = 0.483 and on 12 degrees of freedom

SEM Example. Extract coefficients

```

    graph LR
      A[Latitude] --> B[Degree days]
      B --> C[Date]
      C --> D[Growth]
      D --> E[Survival]
  
```

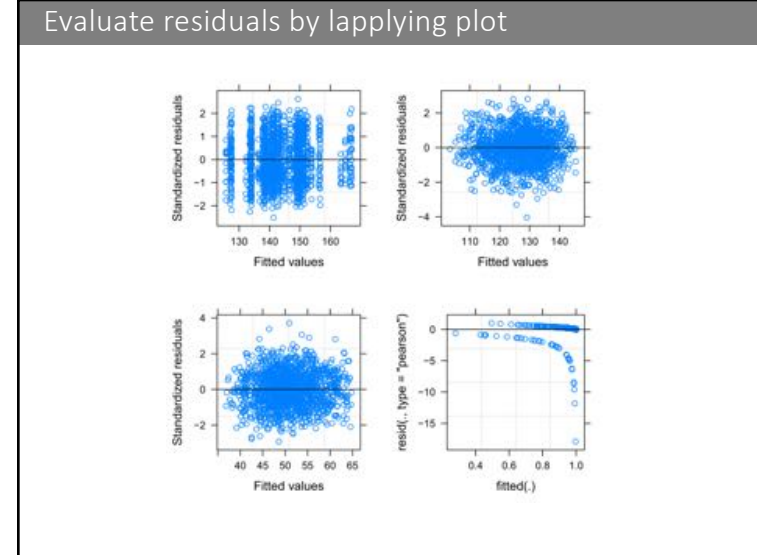
Coefficients:

| Response | Predictor | Estimate | Std.Error | DF | Crit.Value | P.Value | Std.Estimate |
|----------|-----------|----------|-----------|------|------------|---------|--------------|
| DD | lat | -0.8355 | 0.1194 | 18 | -6.9960 | 0 | -0.6877 *** |
| Date | DD | -0.4976 | 0.0049 | 1330 | -100.8757 | 0 | -0.6281 *** |
| Growth | Date | 0.3007 | 0.0266 | 1330 | 11.2917 | 0 | 0.3824 *** |
| Live | Growth | 0.3479 | 0.0584 | 1431 | 9.9552 | 0 | NA *** |

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05

Individual R-squared:

| Response | method | Marginal | Conditional |
|----------|--------|----------|-------------|
| DD | <NA> | 0.49 | 0.70 |
| Date | <NA> | 0.41 | 0.98 |
| Growth | <NA> | 0.11 | 0.84 |
| Live | delta | 0.16 | 0.18 |



SEM Example. For GLMMs, use DHARMA

The figure shows two DHARMA scaled residual plots. The left plot is a QQ plot of residuals, showing observed values on the y-axis (0.0 to 1.0) and expected values on the x-axis (0.0 to 1.0). The right plot is a residual vs. predicted quantile plot, showing standardized residuals on the y-axis (0.0 to 1.0) and predicted values on the x-axis (0.3 to 0.9). Red horizontal lines are drawn at 0.25, 0.5, and 0.75.

```

#residuals for a glmm
library(DHARMA)
sims <- simulateResiduals(shipley.sem[[4]])
plot(sims)
  
```

SEM Example. Populate final model

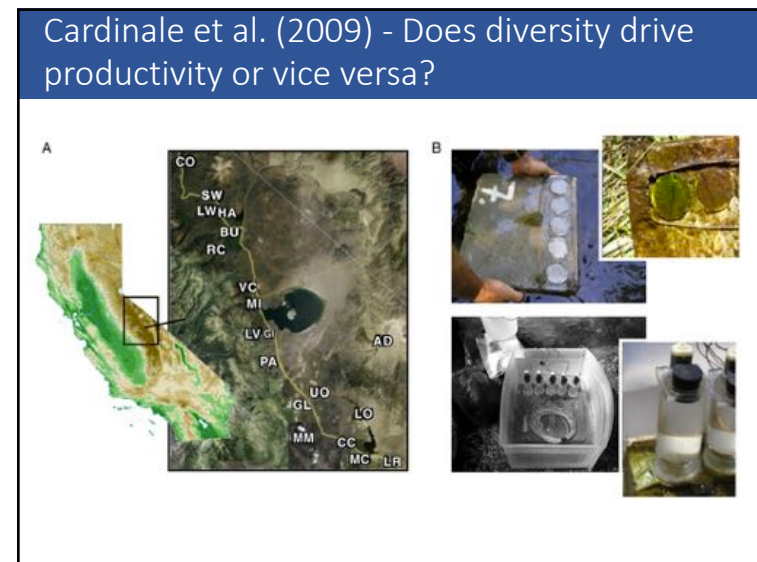
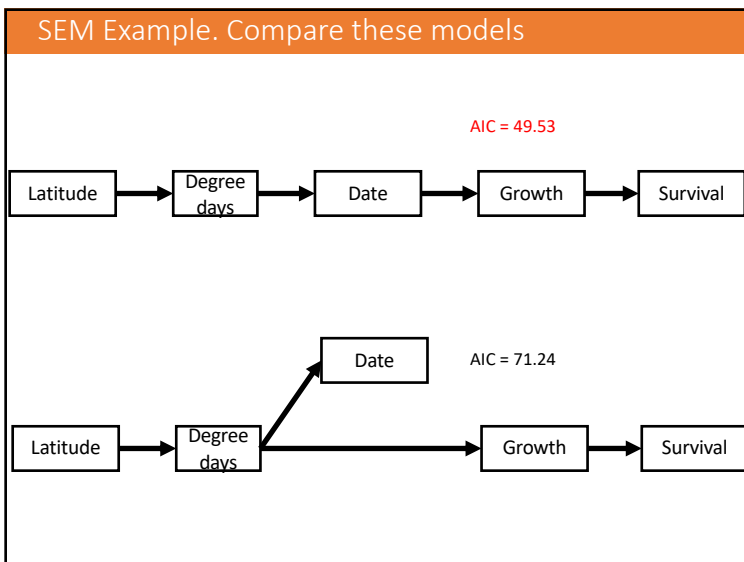
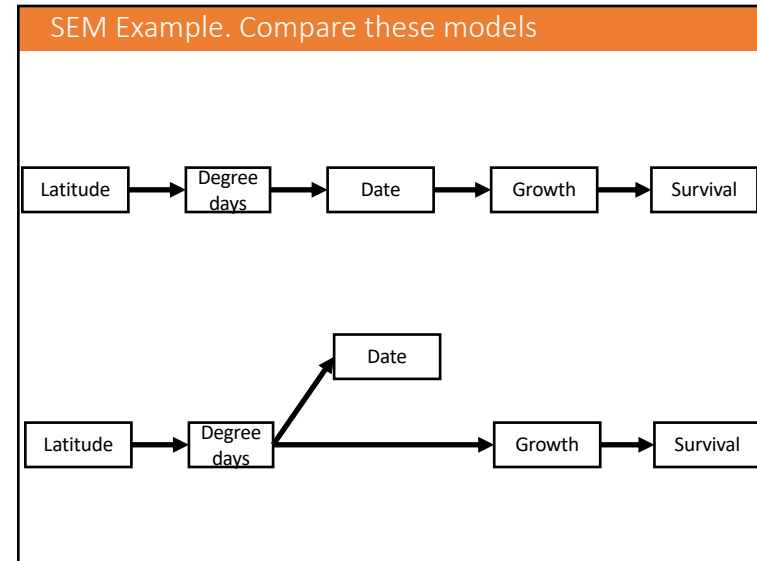
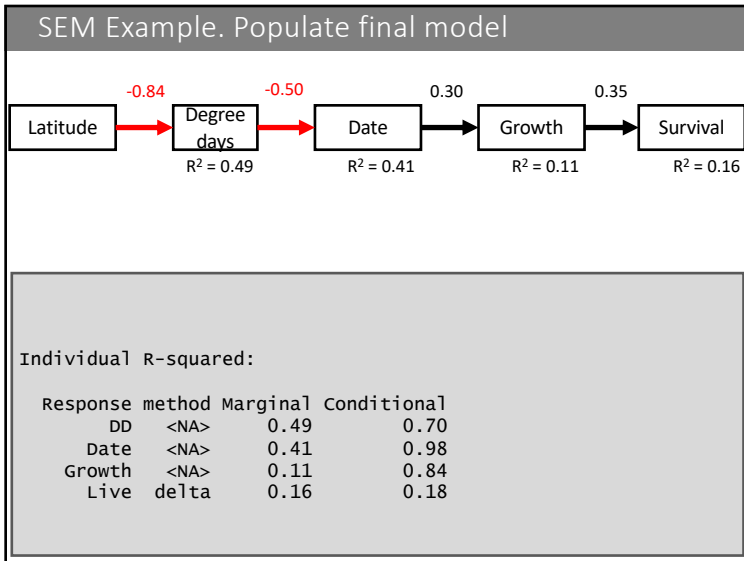
```

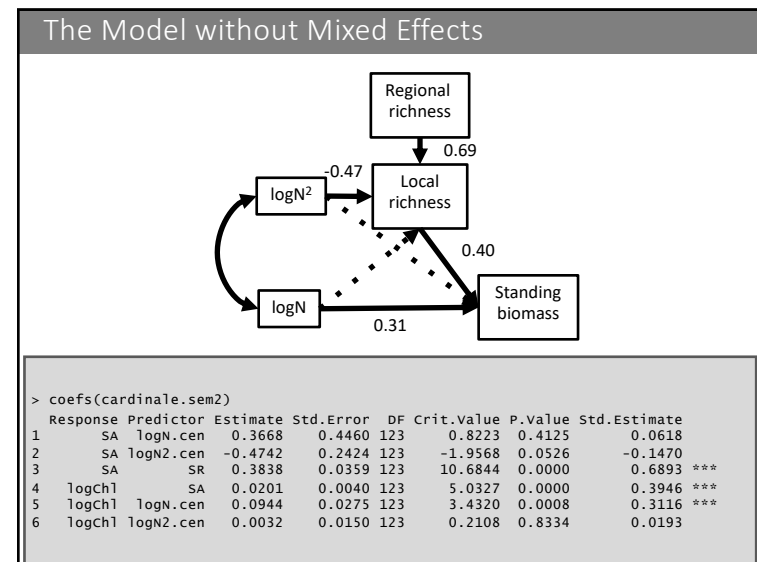
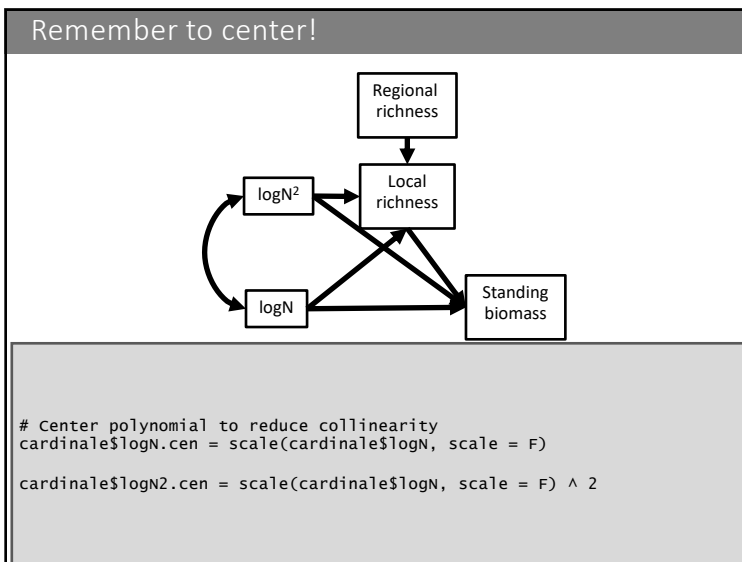
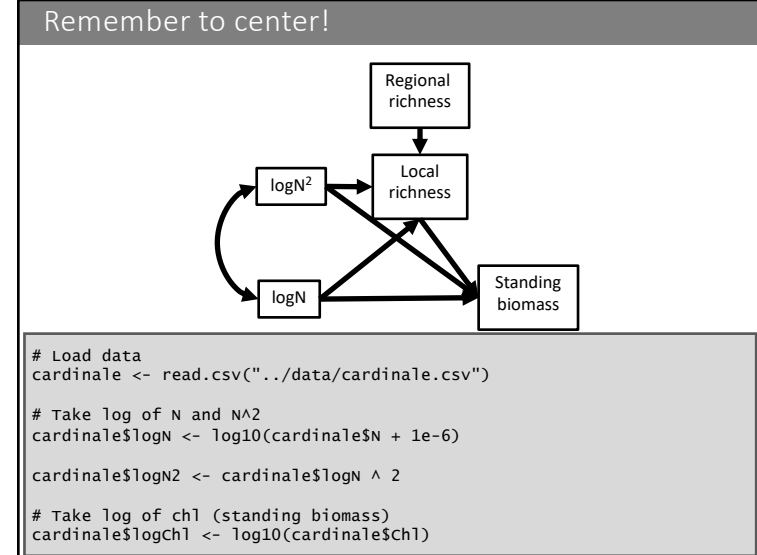
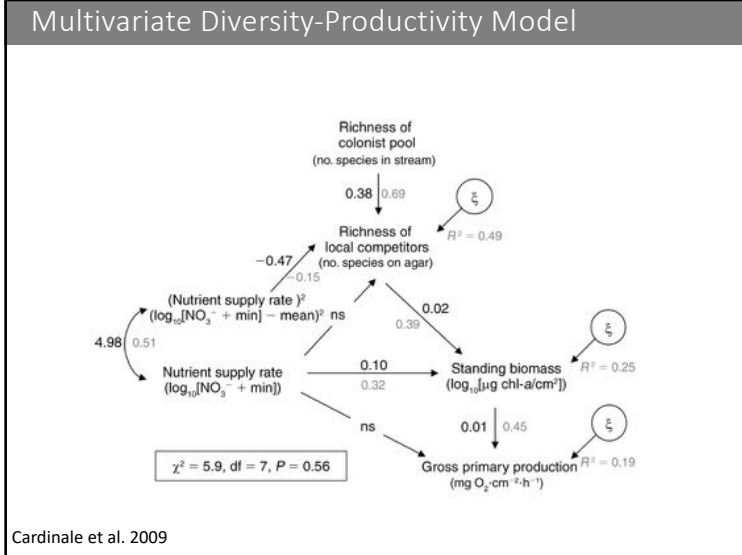
    graph LR
      A[Latitude] -- -0.84 --> B[Degree days]
      B -- -0.50 --> C[Date]
      C -- 0.30 --> D[Growth]
      D -- 0.35 --> E[Survival]
      B --- B_R2["R² = 0.49"]
      C --- C_R2["R² = 0.41"]
      D --- D_R2["R² = 0.11"]
      E --- E_R2["R² = 0.16"]
  
```

Coefficients:

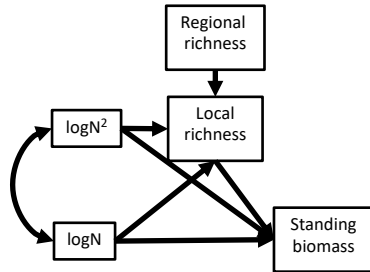
| Response | Predictor | Estimate | Std.Error | DF | Crit.Value | P.Value | Std.Estimate |
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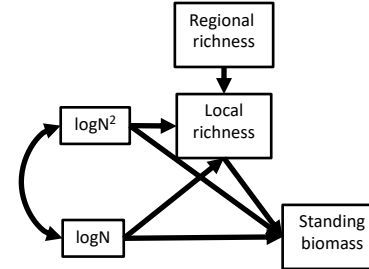


Exercise: Fit with Stream as grouping variable



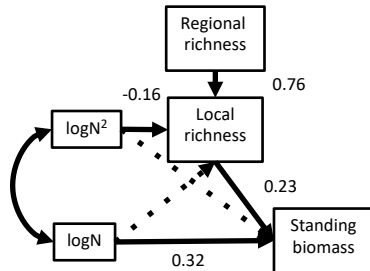
1. What does the model look like?
2. How does it differ from the fixed effects only model?

The Model without Mixed Effects



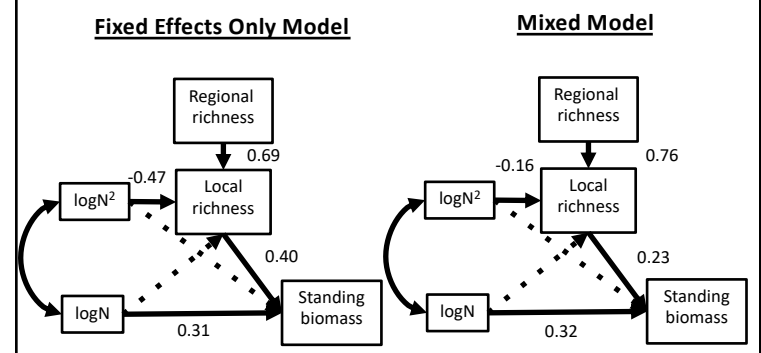
```
# Re-fit SEM using centered predictors
cardinale.mixed <- psem(
  lme(SA ~ logN.cen + logN2.cen + SR,
      random = ~1|Stream, data = cardinale),
  lme(logCh1 ~ SA + logN.cen + logN2.cen,
      random = ~1|Stream, data = cardinale),
  data = cardinale
)
```

The Model without Mixed Effects



```
> coefs(cardinale.mixed)
Response Predictor Estimate Std.Error DF Crit.Value P.Value Std.Estimate
1 SA logN.cen 0.4356 0.3279 105 1.3284 0.1869 0.0734
2 SA logN2.cen -0.5136 0.1783 105 -2.8806 0.0048 -0.1592 **
3 SA SR 0.4260 0.0663 18 6.4251 0.0000 0.7651 ***
4 logCh1 SA 0.0117 0.0050 104 2.3525 0.0205 0.2285 *
5 logCh1 logN.cen 0.0970 0.0223 104 4.3535 0.0000 0.3205 ***
6 logCh1 logN2.cen -0.0022 0.0123 104 -0.1811 0.8566 -0.0136
```

Compare with v. Without Random Effects



Compare R²

```

> rsquared(cardinale.sem2)
Response family link method R.squared
1 SA gaussian identity none 0.4882395
2 logCh1 gaussian identity none 0.2538638

> rsquared(cardinale.mixed)
Response family link method Marginal Conditional
1 SA gaussian identity none 0.5255357 0.7535010
2 logCh1 gaussian identity none 0.1568633 0.4773681
    
```

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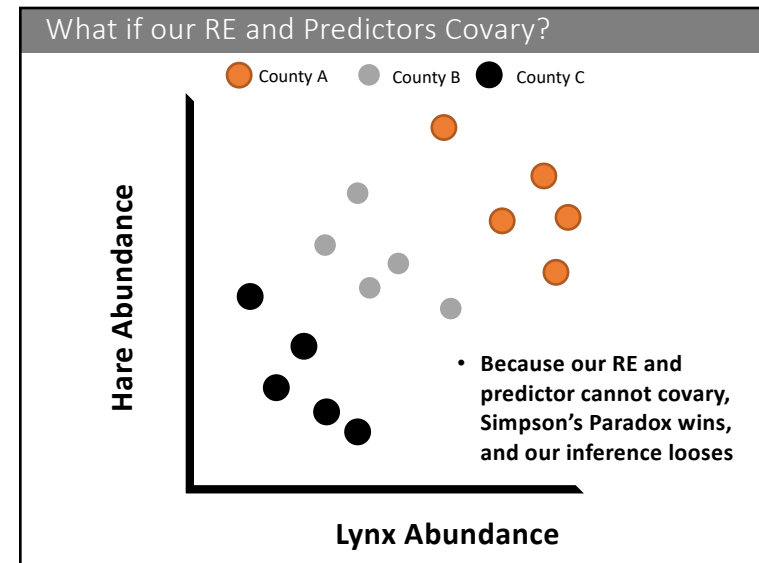
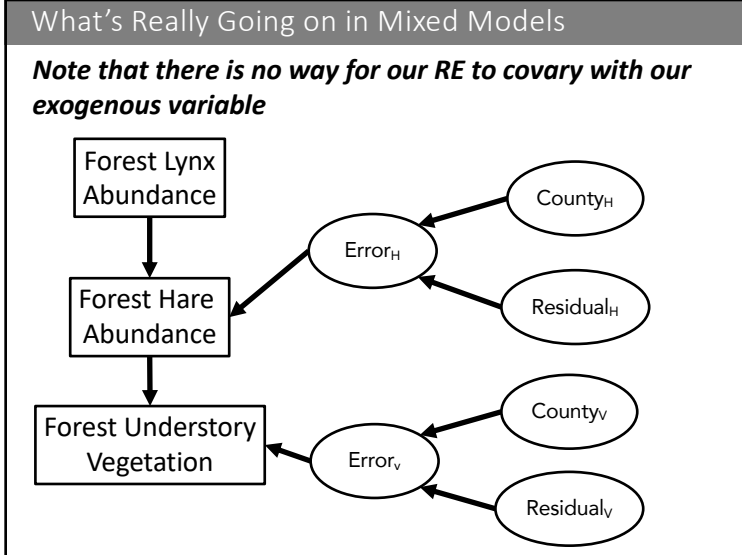
Mixed Models and Graphical Models

Random Effects are latents...

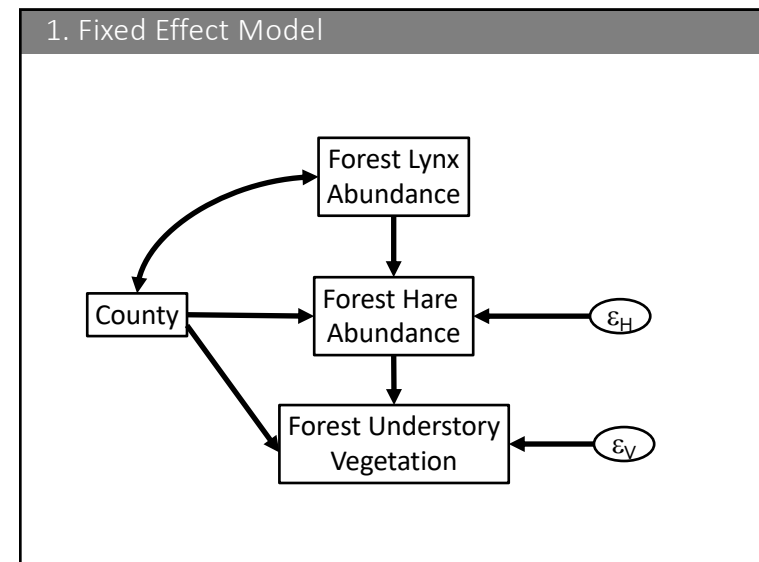
Mixed Models and Graphical Models

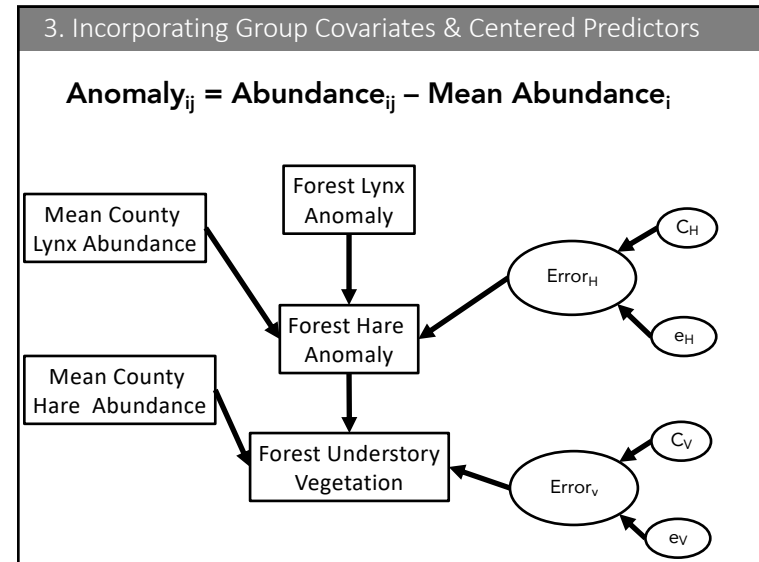
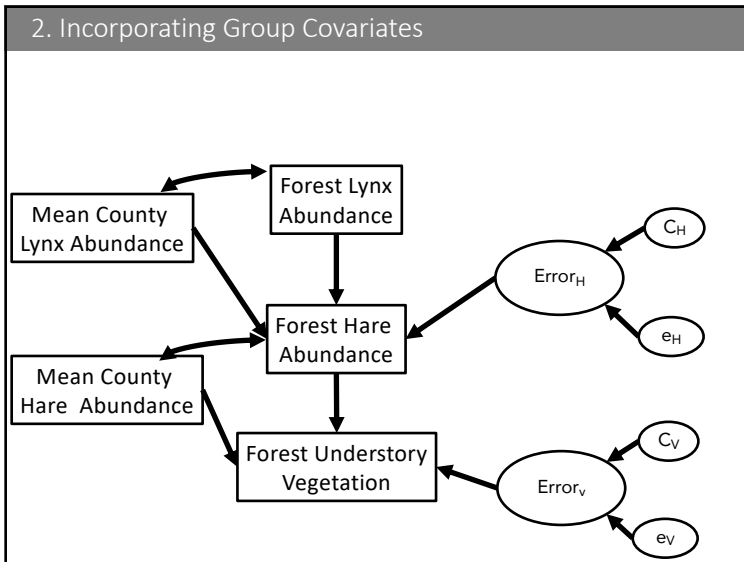
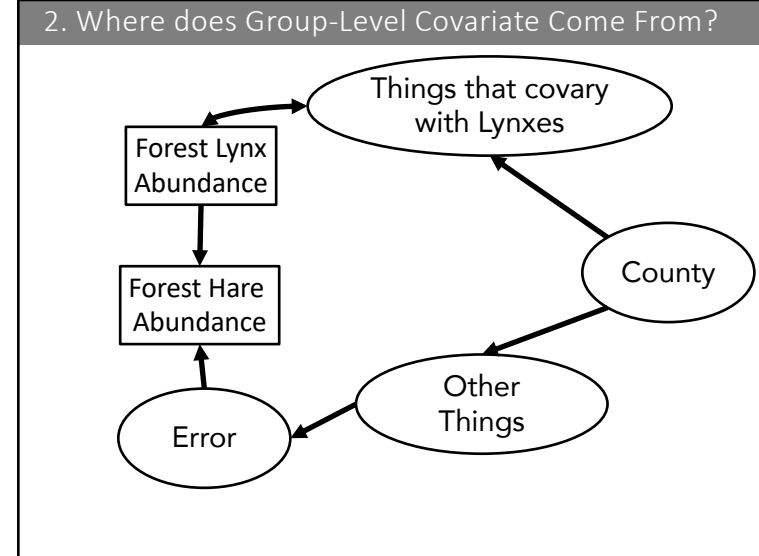
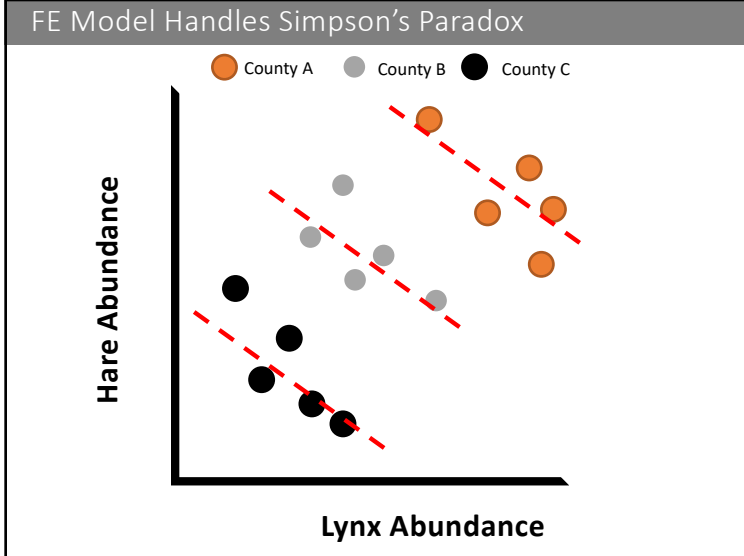
Error is also a latent with mean 0 and some SD

Really, mixed model error is RE variability + Residual variability



- ### Solutions to our RE and Predictors Covarying
1. Have our RE as a fixed effect
 - Can have interaction effects for variable slopes
 - BUT – can cost DF, and open to critique of generalizability
 - BUT – that doesn't matter if you are interested in causal identifiability
 2. Include centered group-level predictor and RE
 - Covariate effect now estimated after controlling for correlation with group level mean
 - Understanding that correlation can be tricky
 - Interpretation of group-level covariate difficult
 3. Include centered group-level predictor, deviation from group level predictor, and RE
 - Correlation broken, so both terms easier to interpret
 - Caution: group-level predictor contaminated by other site-level effects





Are Random Effects Always the Answer?

- No!
- We need to be careful that we are not opening a new back door by relying on random effects
- But, through careful consideration of model structure, we can hold that back door shut, and then some!

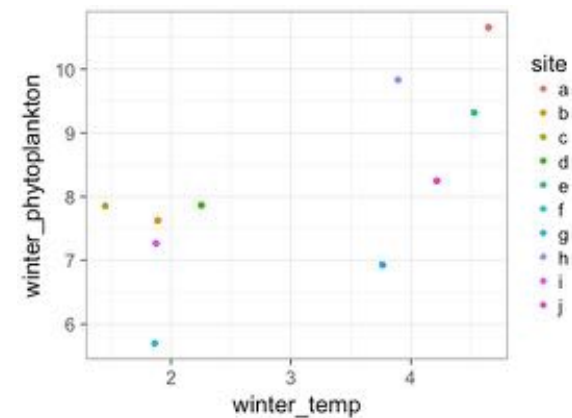
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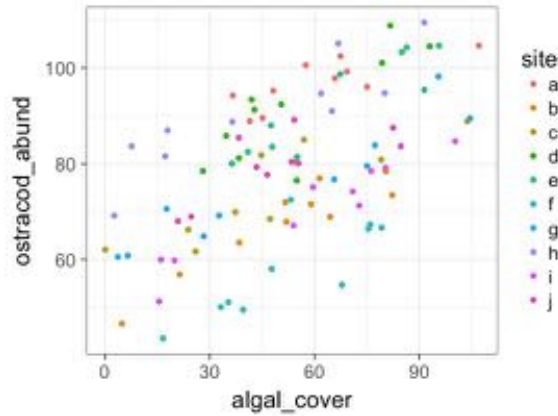
Filter Feeding Ostracods Living In Algae



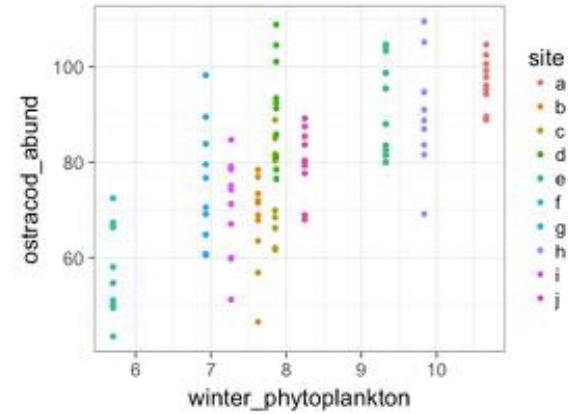
Site-Level Environmental Relationships



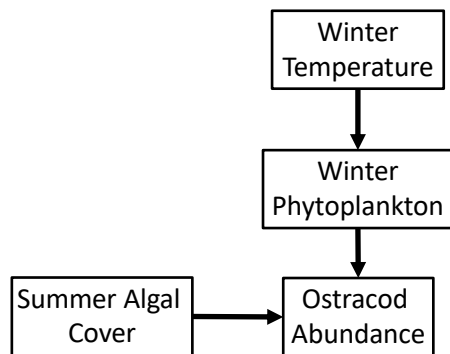
Plot-Level Biotic Relationships



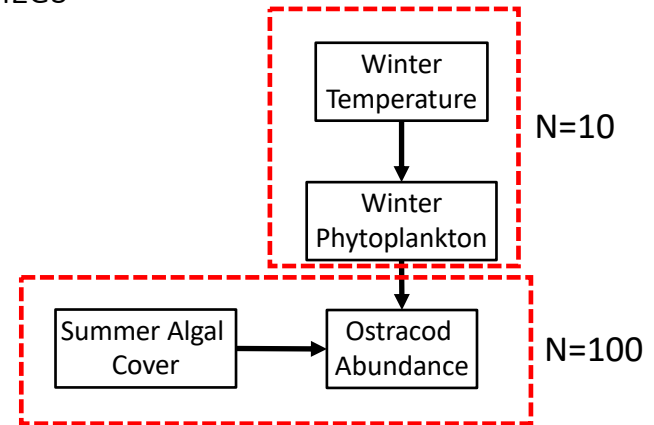
But...site-level drivers of local phenomena



Our Model: What is the basis set?



The problem: Variable Sample Sizes

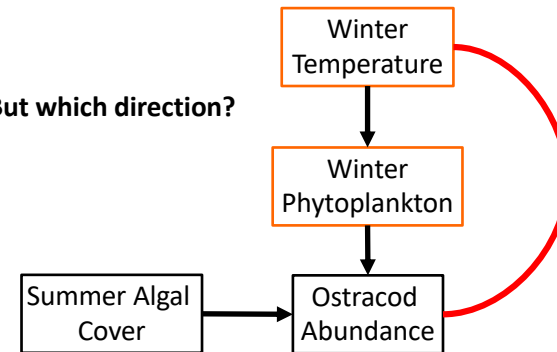


Quandaries with hierarchical SEM

- What **is** our sample size?
 - To some extent solved by hierarchical linear models
 - But, different model components will have different n – and hence different power
- How do we evaluate the basis set?
 - Trickier...but, we can manage!

Our Basis Set

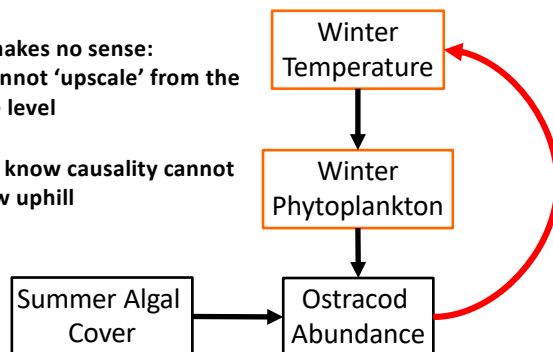
But which direction?



Basis Solutions

This makes no sense:

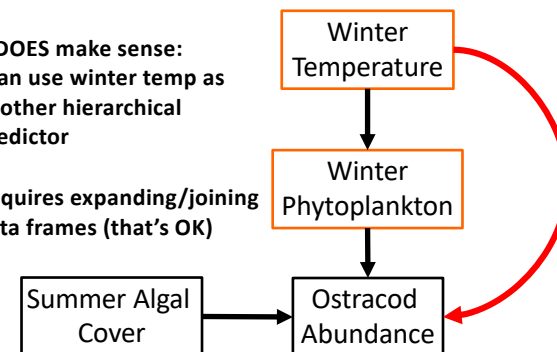
- Cannot 'upscale' from the site level
- We know causality cannot flow uphill



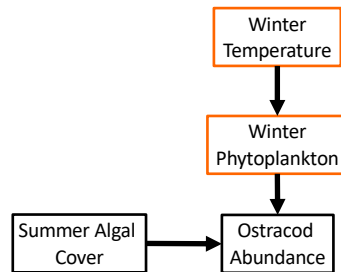
Basis Solutions

This DOES make sense:

- Can use winter temp as another hierarchical predictor
- Requires expanding/joining data frames (that's OK)



Our Model: Two Pieces (for now)



```

ostra_site <- read.csv("../data/ostracod_sitelevel.csv")
ostra_plot <- read.csv("../data/ostracod_plotlevel.csv")

```

Our Model: Two Pieces (for now)

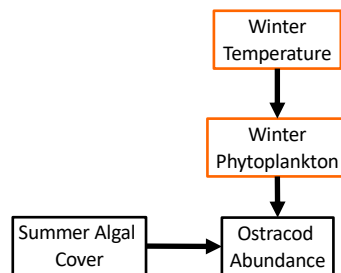
```

#site level
ostra_site_model <- psem(
  lm(winter_phytoplankton ~ winter_temp, data = ostra_site),
  data = ostra_site
)

#plot level
ostra_plot_model <- psem(
  lme(ostracod_abund ~ algal_cover + winter_phytoplankton,
    random = ~1|site, data = ostra_plot),
  data = ostra_plot
)

```

To get C, sum up C from submodels, and get hierarchical C

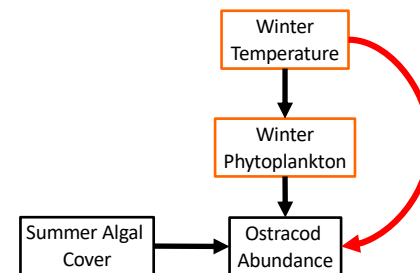


```

> fisherC(ostra_site_model)
Fisher.C df P.value
1 0 0 1
2 > fisherC(ostra_plot_model)
Fisher.C df P.value
1 0 0 1

```

To get C, sum up C from submodels, and get hierarchical C

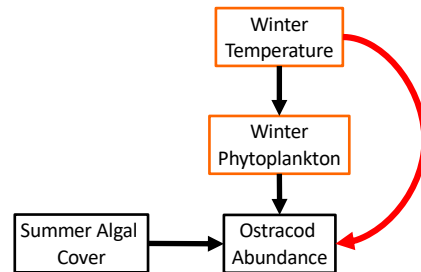


```

basis_mod <- lme(ostracod_abund ~ algal_cover +
  winter_phytoplankton + winter_temp,
  random = ~1|site, data = ostra_plot)

```

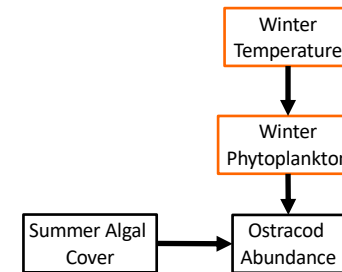
To get C, sum up C from submodels, and get hierarchical C



Fixed effects: ostracod_abund ~ algal_cover + winter_phytoplankton + winter_temp

| | Value | Std.Error | DF | t-value | p-value |
|----------------------|----------|-----------|----|-----------|---------|
| (Intercept) | 5.677315 | 13.167862 | 89 | 0.431149 | 0.6674 |
| algal_cover | 0.324035 | 0.019371 | 89 | 16.728113 | 0.0000 |
| winter_phytoplankton | 6.508925 | 2.080789 | 7 | 3.128105 | 0.0167 |
| winter_temp | 1.210299 | 2.385633 | 7 | 0.507328 | 0.6275 |

To get C, sum up C from submodels, and get hierarchical C



```

> fish_c <- 0 + 0 + -2*log(0.6275)
> 1 - pchisq(fish_c, df = 1)
[1] 0.3343377
  
```

Hierarchical Models in SEM

- This is a new and fast developing area
 - Additional methods in next version of lavaan, too
- In essence, everything is the same...
- Except we need to think carefully about what is the correct test of conditional independence
- Otherwise, we use conventional HLMs, as in a univariate sense